

(2S*,3S*,4R*,5R*)-3,4,5-Trihydroxy-6-(hydroxymethyl)-3,4,5,6-tetrahydro-2H-pyran-2-yl benzoate**Xiang-Jiu He,^a Emil Lobkovsky^b and Rui Hai Liu^{a*}**^aDepartment of Food Science and Institute of Comparative and Environmental Toxicology, Cornell University, Ithaca, NY 14853, USA, and^bDepartment of Chemistry and Chemical Biology, Cornell University, Ithaca, NY 14853, USA

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Key indicators

Single-crystal X-ray study

T = 173 K

Mean $\sigma(\text{C}-\text{C}) = 0.001 \text{ \AA}$

Disorder in main residue

R factor = 0.032

wR factor = 0.092

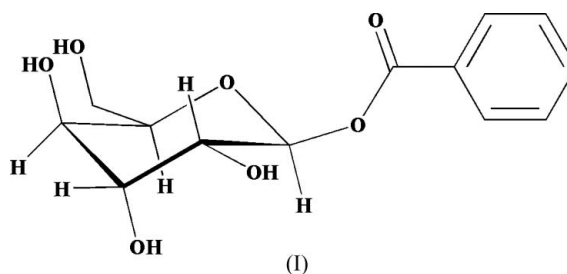
Data-to-parameter ratio = 15.5

For details of how these key indicators were automatically derived from the article, see <http://journals.iucr.org/e>.

The title compound, $\text{C}_{13}\text{H}_{16}\text{O}_7$, was extracted from fresh cranberries (*Vaccinium macrocarpon*). In the crystal structure, molecules are linked into a two-dimensional framework *via* $\text{O}-\text{H}\cdots\text{O}$ hydrogen bonds.

Comment

Cranberries (*Vaccinium macrocarpon*), a native fruit of North America, have attracted public attention due to potential health benefits. Recently, cranberries have been found to be rich in phenolics (Sun *et al.*, 2002), to prevent bacterial adhesion in urinary tract infections of *E. coli* and stomach ulcers (Howell *et al.*, 1998; Foo *et al.*, 2000; Burger *et al.*, 2000), to exhibit *in vitro* anticancer activity (Sun *et al.*, 2002; Bomser *et al.*, 1996) and to protect against lipoprotein oxidation (Chu & Liu, 2005; Wilson *et al.*, 1998). The attractive bright red appearance and distinctive flavor of cranberries is recognized as a concentrated source of dietary flavonoids (Sun *et al.*, 2002), including anthocyanins (Zapsalis & Francis, 1965), proanthocyanidins (Foo & Porter, 1980; Hale *et al.*, 1986) and flavonol glycosides (Puski & Francis, 1967), as well as various phenolic acids (Schmid, 1977).



In our continuing efforts to seek bioactive components from fruits, vegetables and other natural products, bioactivity-guided fractionation of cranberries was used to determine the identity of the bioactive compounds from cranberries, which inhibit tumor cell growth and may play a role in cancer prevention and therapy (He *et al.*, 2005). The title compound, (I), was isolated from the active fraction of cranberry extracts. This compound may be responsible for the antimicrobial activity of cranberries.

A view of the title molecule is shown in Fig. 1. In the crystal structure, intermolecular $\text{O}-\text{H}\cdots\text{O}$ hydrogen bonds connect molecules into a two-dimensional framework (Table 1).

Experimental

Fresh cranberries of Stevens cultivar (7.0 kg) were homogenized five times with chilled 80% acetone (1:2 *w/v*) using a chilled Waring

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blender (Sun *et al.*, 2002). The samples were then homogenized for an additional 3 min using a Polytron homogenizer. The homogenates were filtered and the filtrate was evaporated under vacuum at 318 K until approximately 90% of the filtrate had been evaporated. The residue was then recovered with 4000 ml water and extracted with the same volume of ethyl acetate three times, and then water-saturated *n*-butanol three times. The butanol fraction (204.0 g) was subjected to Diaion HP-20 column chromatography (55 × 550 mm) and eluted with 3000 ml water, 30% methanol (*v/v*), 50% methanol (*v/v*) and methanol. The 30% methanol elution (16.50 g) was further isolated by a silica-gel column (230–400 mesh, 75 × 235 mm) and eluted with dichloromethane–methanol with a gradually increasing proportion of methanol. The title compound (I) (605.2 mg) was obtained from dichloromethane–methanol elution (10:1, *v/v*) as colorless crystals. Spectroscopic analysis: ¹H NMR (400 MHz, DMSO-*d*₆, δ, p.p.m.): 8.04 (2H, *dt*, *J* = 8.7 and 1.5 Hz), 7.70 (H, *td*, *J* = 7.5 and 1.5 Hz), 7.56 (2H, *td*, *J* = 7.5 and 1.5 Hz), 5.61 (H, *d*, *J* = 7.5 Hz), 5.45 (H, *d*, *J* = 5.1 Hz), 5.20 (H, *d*, *J* = 4.8 Hz), 5.08 (H, *d*, *J* = 5.4 Hz), 4.62 (H, *t*, *J* = 6.0 Hz), 3.69 (H, *ddd*, *J* = 12.0, 5.4 and 1.5 Hz), 3.49 (H, *p*, *J* = 6.0 Hz), 3.37–3.16 (4H, *m*); ¹³C NMR (400 MHz, DMSO-*d*₆, δ, p.p.m.): 164.8 (C), 133.9 (C), 129 (2CH), 129.2 (CH), 128.9 (2CH), 95.1 (CH), 78.0 (CH), 76.4 (CH), 72.6 (CH), 69.5 (CH), and 60.0 (CH₂).

Crystal data

C₁₃H₁₆O₇ Mo Kα radiation
M_r = 284.26 Cell parameters from 8636 reflections
 Orthorhombic, *P*2₁2₁2₁ θ = 2.7–37.4°
a = 5.8022 (2) Å μ = 0.12 mm⁻¹
b = 7.6865 (3) Å *T* = 173 (2) K
c = 28.3221 (12) Å Block, colorless
V = 1263.13 (8) Å³ 0.70 × 0.60 × 0.50 mm
Z = 4
D_x = 1.495 Mg m⁻³

Data collection

Bruker X8 APEX-II diffractometer 3448 reflections with *I* > 2σ(*I*)
 φ and ω scans *R*_{int} = 0.024
 Absorption correction: multi-scan θ_{max} = 37.8°
 (SADABS; Sheldrick, 1996) *h* = -10 → 6
*T*_{min} = 0.919, *T*_{max} = 0.941 *k* = -13 → 7
 15473 measured reflections *l* = -48 → 39
 3768 independent reflections

Refinement

Refinement on *F*² H atoms treated by a mixture of
R[*F*² > 2σ(*F*²)] = 0.032 independent and constrained
wR(*F*²) = 0.092 refinement
S = 1.06 *w* = 1/[σ²(*F*_o²) + (0.0678*P*)²]
 3768 reflections where *P* = (*F*_o² + 2*F*_c²)/3
 243 parameters (Δ/σ)_{max} = 0.001
 Δρ_{max} = 0.41 e Å⁻³
 Δρ_{min} = -0.21 e Å⁻³

Table 1

Hydrogen-bond geometry (Å, °).

<i>D</i> –H... <i>A</i>	<i>D</i> –H	H... <i>A</i>	<i>D</i> ... <i>A</i>	<i>D</i> –H... <i>A</i>
O1–H1O...O7 ⁱ	0.97 (3)	1.81 (3)	2.7607 (10)	166 (2)
O3–H3O...O2 ⁱⁱ	0.82 (2)	1.88 (2)	2.6774 (10)	166.1 (16)
O4–H4A...O1 ⁱⁱⁱ	0.84	2.19	3.0121 (12)	167
O4A–H4AA...O7 ⁱⁱⁱ	0.84	2.19	2.953 (11)	151

Symmetry codes: (i) *x* + 1, *y*, *z*; (ii) *-x* + 1, *y* + ½, *-z* + ¾; (iii) *x*, *y* + 1, *z*.

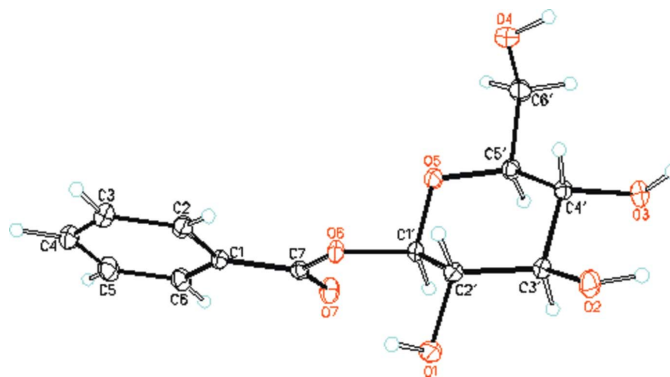


Figure 1 Molecular structure of (I), showing 40% probability displacement ellipsoids and H atoms as spheres of arbitrary radii. Only the major disorder component is shown.

In the absence of significant anomalous dispersion effects, Friedel pairs were merged before the refinement and the assignment of the absolute stereochemistry is arbitrary. The methylhydroxy group containing atom O4 is disordered over two sites with refined occupancies of 0.889 (3) and 0.111 (3) for the major and minor components, respectively. The disorder corresponds to a rotation about the C5'–C6' bond. The disordered H atoms were included in calculated positions, with C–H = 0.99 Å and *U*_{iso}(H) = 1.2*U*_{eq}(C), and O–H = 0.84 Å and *U*_{iso}(H) = 1.5*U*_{eq}(O). All other H atoms were refined isotropically. The C–H distances are in the range 0.936 (15)–1.026 (15) Å.

Data collection: APEX2 (Bruker, 2004); cell refinement: SAINT-Plus (Bruker, 2003); data reduction: SAINT-Plus; program(s) used to solve structure: SHELXTL (Bruker, 1999); program(s) used to refine structure: SHELXTL; molecular graphics: SHELXTL; software used to prepare material for publication: SHELXTL.

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